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Rearrangements of 14-Mesyloxy-*ent*-Beyer-15-enes

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Abstract: Rearrangement of *ent*-7 α ,18-diacetoxy-14 β -mesyloxybeyer-15-ene (4) under several conditions gave 12(13 \rightarrow 14)-*abeo* compounds (5 or 7), on some occasions in high yield, which evolved to 12(13 \rightarrow 14),15(8 \rightarrow 7)-*diabeo* compounds in acidic medium. Structures of rearranged products were established principally with the aid of INADEQUATE and other 2D-nmr experiments.

INTRODUCTION

Rearrangements of tetracyclic diterpenes have normally been achieved shifting leaving groups from C-12 of *ent*-kaurane¹ or *ent*-beyerane², from C-16 of *ent*-kaurane³ and *ent*-beyerane^{2a,4} and from C-13 of *ent*-atisane skeletons^{1a}. On a few occasions the rearrangement processes were initiated by an electronic deficiency on C-15⁵ of beyerane or on C-14 of *ent*-kaurane compounds⁶.

We previously reported the influence of another functional group on the cycles to be rearranged in the solvolysis process, of 12,17-difunctionalized *ent*-beyeranes⁷ compounds, and demonstrated the influence of some groups at C-14⁸ on processes of rearrangement initiated on C-16 of *ent*-15 α ,16 α -epoxybeyerane compounds. We now report the first rearrangement initiated at C-14 of *ent*-beyer-15-ene compounds.

RESULTS AND DISCUSSION

ent-14 β ,18-Dihydroxy-7 α -acetoxybeyer-15-ene (7-acetylpusillatriol, 1) is a natural product isolated from *Sideritis* ⁹. Acetylation of 1 under mild conditions¹⁰ gave the diacetate 2 (75%) and triacetate 3 (20%). Mesylation of compound 2 gave mesylate 4 (95%), which was treated under several conditions to give, preferentially, rearranged products. Thus, treatment of 4 with LiAlH₄ under the normal conditions of demesylation gave principally product 5 (48%) and pusillatriol 6 (35%)¹¹. Nmr spectra of product 5 (see Table 1 and experimental part) clearly indicated the presence of two unsaturated groups, one of which was an

exomethylene group, which was not possible in an *ent*-beyerene skeleton. Hence a rearrangement process might be involved, probably initiated at C-14 by the mesyloxy group. Product 4 was treated under AcOH/AcOK solvolysis conditions to demonstrate this possible rearrangement. In this case (pathway A in Figure 1) product 7 (with the same type of skeleton as product 5) was obtained in high yield (95%). A similar result (90%) was obtained when mesylate 4 was treated with Zn/EtOH (pathway B in Figure 1). Connectivity between the carbon atoms in rearranged product 7 was determined by INADEQUATE experiments (see Figure 2) done with a refocalization time of 0.0065 seconds, calculated for a value of 38.5 Hz for the C/C coupling constant, which is an average value for saturated carbons. Thus, as can be observed in Figure 2, all the connectivities could be determined with the exception of C-15/C-16 and C-13/C-16, which have estimated coupling constants of around 76 Hz¹².

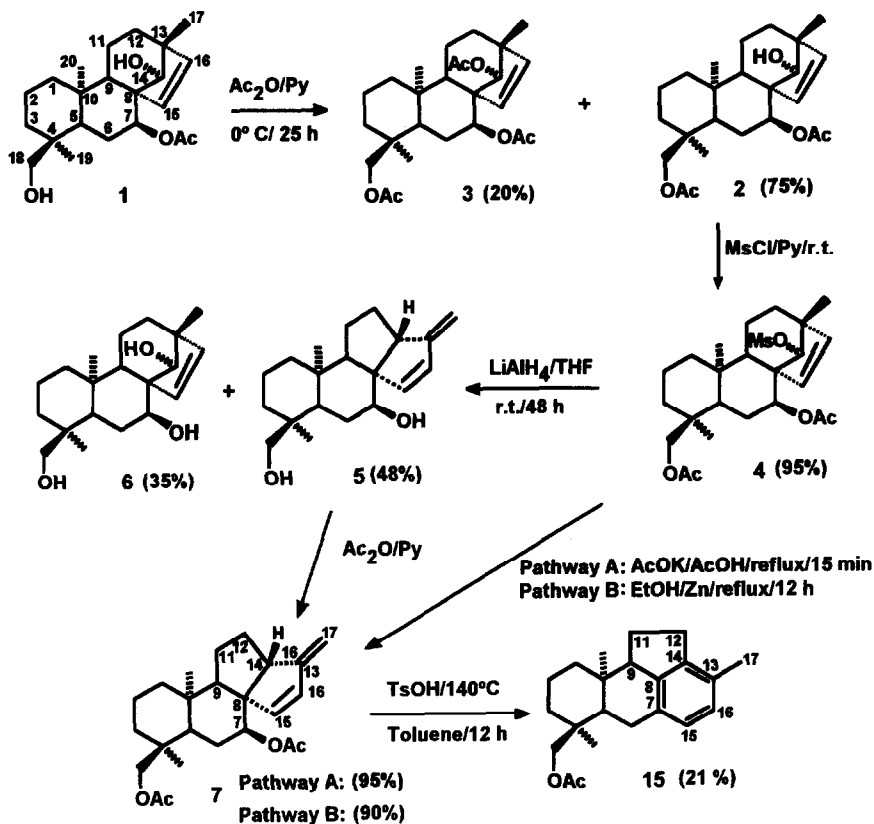


FIGURE 1

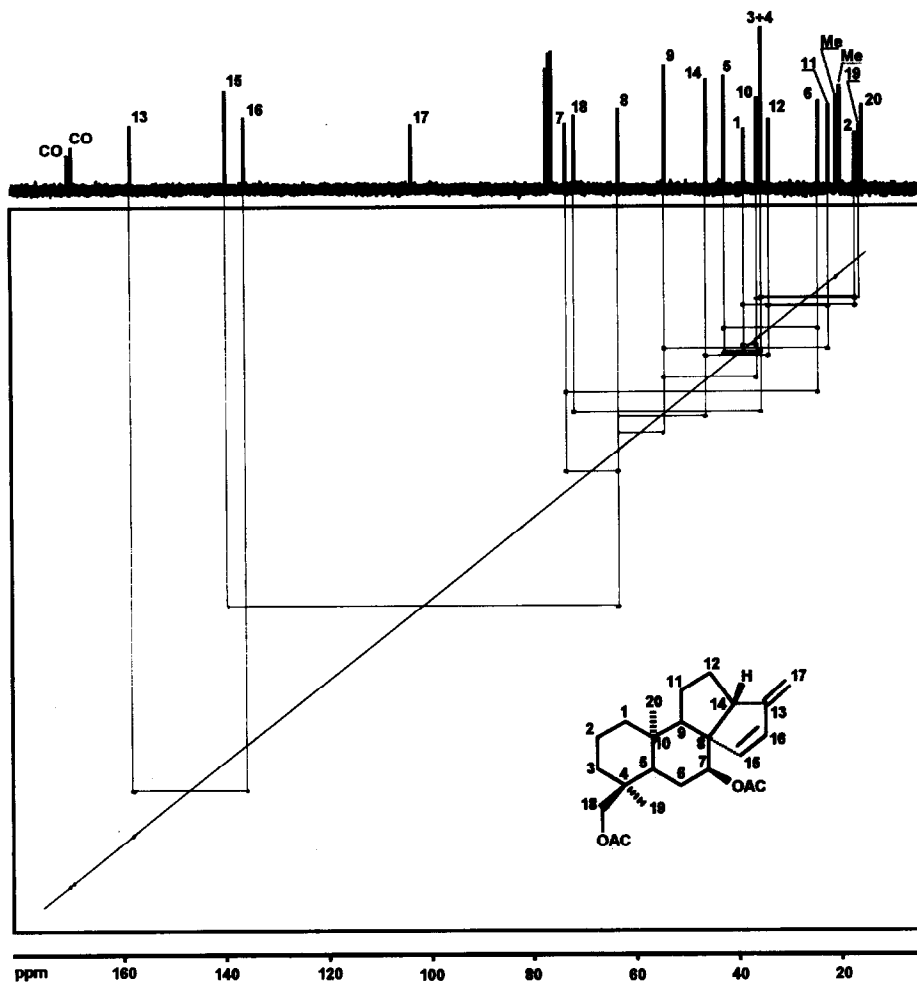
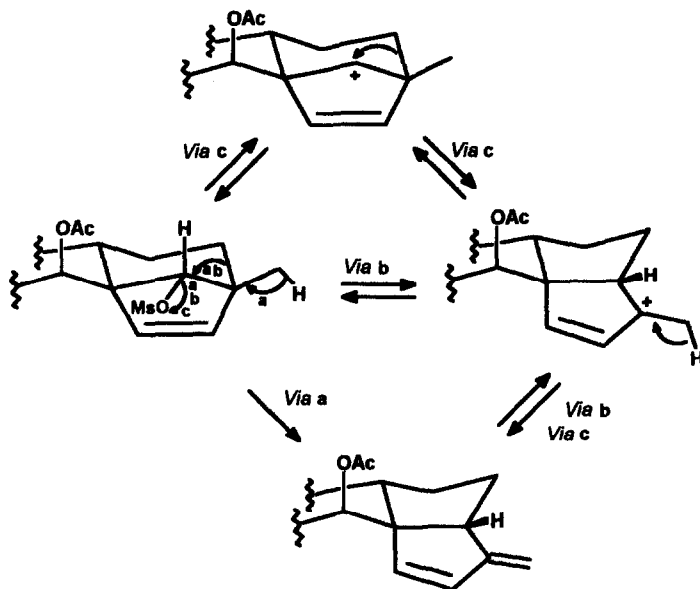


FIGURE 2

After these assignments, heteronuclear C/H correlation and proton-proton homonuclear correlation (COSY90) were done to assign the proton signals of product 7. This experiment allowed us to assign the skeleton indicated in structure 7, and its stereochemistry was determined by phase sensitive NOESY, after assignment of the protons by heteronuclear C/H correlation. The dipolar correlation signal appeared between 3H-20 (δ 0.91) and the protons at C-15 (δ 6.00). Hence, the stereochemistry of the new cycles at C-8 was maintained in a rearrangement process that occurred by migration of bonds C-12/C-13 to C-12/C-14, as



SCHEME 1

shown in Scheme 1. Taking into account the *trans*-periplanar arrangement of the mesyloxy group and the bond, the process can be completely concerted (*pathway a*), although an allylic carbocation may also have been formed (*pathway b*). The configuration at C-14 can be fixed in route *a* or *b*. The calculated energy for the 14 β -H configuration was 55.28 kcal/mol, and the calculated coupling constants ($J_1=9.04$ Hz, $J_2=1.39$ Hz) were in agreement with the experimental values ($J=10.79$ Hz, *bd*), although the program for the calculations

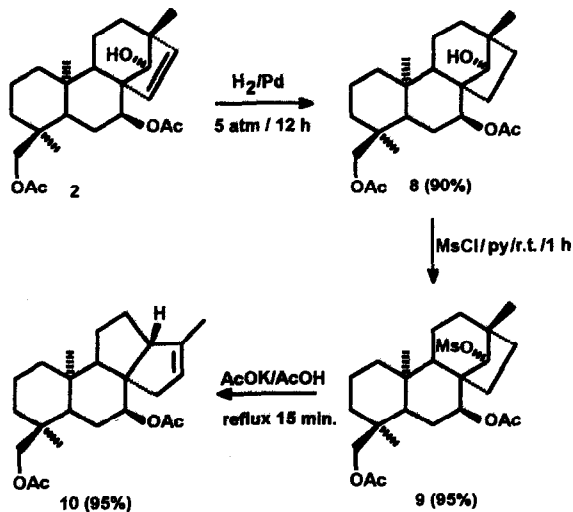
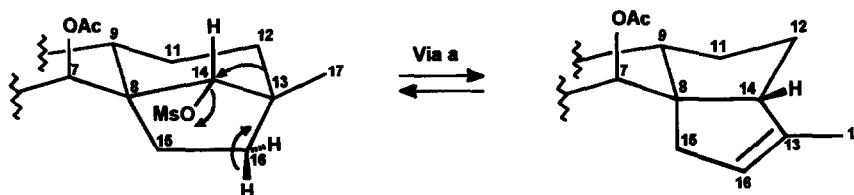


FIGURE 3

failed for conjugated system¹³. The other possible configuration at C-14 was not possible (133.8 kcal/mol). The high yield of the rearrangement (still 95%) seemed to indicate that the main pathway was *a* or *b*.

We also achieved the rearrangement of saturated compound 9 (Figure 3) under the same conditions as indicated for unsaturated product 4 to give, also in high yield, a rearranged product 10 with the same skeleton as that described for product 7, although in this case the carbocation at C-13 was stabilised by loss of a proton from C-16 to give an *endo*-cyclic double bond (see Scheme 2, calculated energy was 47.18 kcal/mol for *endo*-cyclic double bond compound, and 50.97 kcal/mol for the *exo*-cyclic double bond product).



SCHEME 2

We also did 2D-nmr experiments in addition to monodimensional pmr and cmr (see Table 1) to determine the stereochemistry of rearranged product 10. After assignation of the largest proton signals by C/H correlation, we detected (NOESY) dipolar correlation between the geminal proton of C-7 and of the allylic protons of C-15 (δ 2.48). The other allylic protons of C-15 (δ 1.68) showed a dipolar correlation with the methyl group of C-20 (δ 0.83). The configuration at C-14 was also defined by the rearrangement process. Calculated energy for product 10 was 54.45 kcal/mol (rather than 85.72 for its C-14 epimer) and the calculated coupling constants ($J_1=10.20$ Hz, $J_2=2.90$ Hz) were in agreement with the experimental values ($J=10.40$ Hz, bd), rather than calculated values ($J_1=12.24$ Hz, $J_2=4.94$ Hz) for its 14 β -epimer.

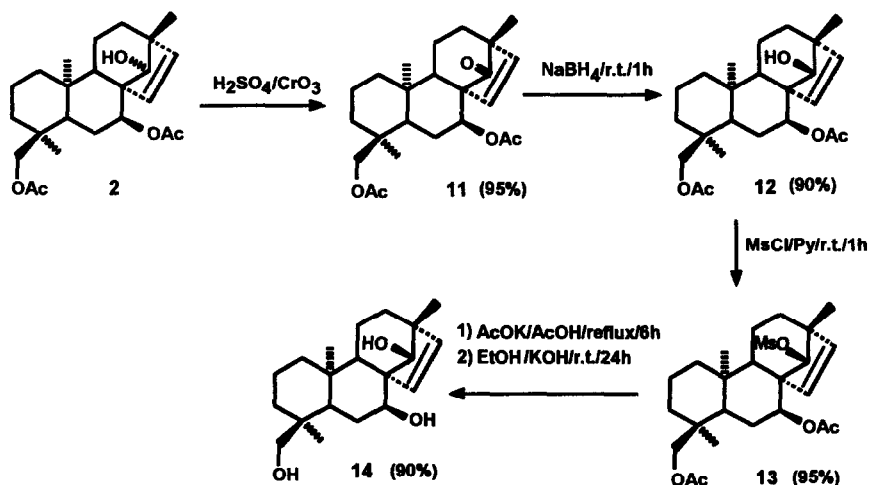
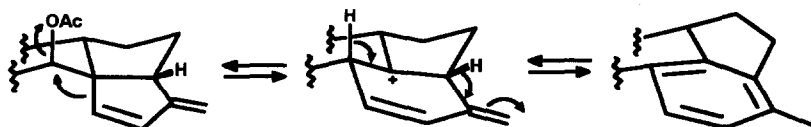


FIGURE 4

The result of the rearrangement of this saturated product **8** seems to indicate that these rearrangements are not dependent of the formation of an allylic carbocation, hence pathway *a*, with participation of a proton at C-17 in the case of product **4** or at C-16 in the case of product **9**, was preferred.

We also obtained the epimer mesylate compound at C-14 of starting product **13** to confirm the type of these rearrangements. Oxidation of diacetate compound **2** (see Figure 4) gave ketone compound **11** (95%), which was treated with NaBH₄ to give compound **12** (90%). Product **12** was mesylated as usual (see experimental part) to give product **13** (93%). Acetolysis of product **13**, inclusive under higher energetic



SCHEME 3

conditions than those indicated above for mesylate **4**, produced only a inseparable mixture of *ent*-14 α -acetoxy or *ent*-14 α -hydroxy derivatives, which was saponified to give *ent*-7 α ,14 α ,18-trihydroxybeyer-15-ene (**14**, 90%). These reactions ruled out pathway *c* for the rearrangement of products **4** and **9** (see Scheme 1). When we treated rearranged product **7** and **10** with acid, product **7** evolved toward an aromatic product (**15**, 21 %) with participation of the acetoxy group at C-7. Acid treatment of product **10** did not lead to any rearrangement (see Scheme 3).

TABLE 1

C	4	5	6	7	8	9	10	12	13	14	15
1	38.3	39.6	38.7	39.4	39.3	39.0	39.1	38.5	38.5	38.6	39.1
2	17.6	17.9	18.0	17.6	17.8	17.5	17.6	17.7	17.6	18.0	17.6
3	35.2	35.5	35.0	36.0	36.5	35.1	36.1	35.7	35.5	35.2	36.1
4	36.1	37.1	37.0	35.9	35.1	36.1	35.9	36.8	37.2	36.8	36.9
5	41.4	46.5	47.7	43.3	45.4	41.3	43.6	42.0	41.9	39.7	43.6
6	23.7	27.1	26.2	24.9	25.0	23.4	24.1	25.1	25.3	27.6	24.1
7	70.9	71.6	68.8	73.8	130.4	74.8	73.6	78.4	74.9	75.2	73.6
8	57.3	65.7	59.0	63.7	142.0	52.4	57.3	51.9	51.9	52.1	57.3
9	47.6	53.8	47.7	54.8	57.0	51.2	55.1	41.0	41.2	38.9	55.1
10	37.6	37.2	37.2	36.9	36.6	38.0	37.1	36.2	36.1	37.2	37.1
11	18.8	23.2	19.2	22.9	26.8	18.8	23.4	18.6	18.5	18.6	23.4
12	31.9	34.5	32.5	34.5	30.2	29.1	30.3	25.6	26.5	25.8	30.3
13	49.0	159.2	49.0	158.4	130.2	43.9	143.0	45.7	45.7	43.3	143.0
14	98.4	40.5	90.8	46.7	141.6	97.7	53.1	84.1	90.1	84.9	53.1
15	129.7	142.2	131.6	140.1	124.9	34.4	39.6	131.0	131.7	132.0	39.6
16	134.7	135.5	135.1	136.4	127.2	37.8	122.1	136.6	134.8	136.2	122.1
17	22.0	103.4	19.3	104.0	18.3	23.6	15.8	21.8	22.6	21.9	15.8
18	72.3	71.2	70.1	72.3	72.5	72.5	72.5	72.6	72.9	70.1	72.5
19	17.8	17.4	18.0	17.0	17.4	17.8	17.2	17.7	17.8	18.2	17.2
20	15.5	16.7	15.4	16.7	14.1	15.8	15.4	15.8	16.1	15.8	15.4
CH ₃	21.0			20.9	20.8	22.2	21.5	21.6	22.1		21.1
CH ₃	21.1			21.1		21.2	21.1	21.1	21.1		21.5
CH ₃	38.7					38.7			40.1		
CO	171.0			170.0	170.9	171.0	171.0	169.7	171.0		171.0
CO	170.9			170.8		171.0	170.4	170.9	171.2		170.5

EXPERIMENTAL

Measurements of NMR spectra (400.13 MHz ^1H and 100.62 MHz ^{13}C) were done in CDCl_3 (which also provided the lock signal) in a Bruker ARX-400 apparatus. The assignments of ^{13}C chemical shifts were done with the aid of distortionless enhancement by polarization transfer (DEPT) using a flip angle of 135° . 2D-nmr experiments: COSY90, NOESY, direct C/H correlation and INADEQUATE, performed with pullprogs from the Bruker library (COSY90, NOESYST, HXCO and INAD respectively). A sample of 130 mg of product 7, in 0.5 mL CDCl_3 in a 5 mm diameter probe was used for INADEQUATE, which was acquired by 512 experiences of 192 scans, with relaxation delay between scans of 2.2 sec, and refocalization time of $1/4J=0.0065$ sec, in a week-end experiment. Infrared spectra were recorded on a Perkin-Elmer mod. 983 G spectrometer or on a Nicolet 20SX FT-IR spectrometer. Mass spectra were determined with CI (methane) or EI (70 eV) in a Hewlett-Packard mod. 5988 A spectrometer. Optical rotations were measured on a Perkin-Elmer 241 polarimeter at 20° . Elemental analyses were performed on a Fisons EA 1108 Elemental Analyzer. Silica gel SDS 60 A CC (40-60 μm) was used for flash chromatography. CH_2Cl_2 or CHCl_3 containing increasing amounts of Me_2CO was used as the eluent. Analytical plates (silica gel, Merck 60 G) were rendered visible by spraying with $\text{H}_2\text{SO}_4/\text{AcOH}$, followed by heating to 120° .

Acetylation of product 1

Product 1 (500 mg) isolated of *Sideritis pusilla* subsp. *flavovirens* according to reference 9 was dissolved in $\text{Ac}_2\text{O}/\text{Py}$ (1:2) with stirring for 25 h at 25°C . The reaction mixture was diluted with water, extracted with CH_2Cl_2 , washed with saturated KHSO_4 and dried with anhydrous Na_2SO_4 . Chromatography over silica gel yielded 376 mg of *ent*-7 α ,18-diacetoxy-14 β -hydroxybeyer-15-ene (2, 75%)¹⁰ and 100 mg of *ent*-7 α ,14 β ,18-triacetoxybeyer-15-ene (3, 20%)¹⁰.

Mesylation of product 2

Methanesulfonyl chloride (MsCl , 0.8 mL) was added to a solution of 450 mg of product 2 dissolved in 1.6 mL pyridine. The reaction mixture was stirred at room temperature for 1 h. Then the reaction mixture was diluted with CH_2Cl_2 , washed with water and with saturated aqueous KHSO_4 , and concentrated in vacuum. Chromatography on a silica gel column yielded 428 mg of *ent*-7 α ,18-diacetoxy-14 β -mesyloxybeyer-15-ene (4, 95%); m.p.: 178°C ; $[\alpha]_{\text{D}} = 29.1^\circ$ (CHCl_3 , c 1); IR_{vmax} (CHCl_3): 1735, 1340, 1248 and 738 cm^{-1} ; ^1H nmr (400 MHz, CDCl_3): δ 5.54 and 5.50 (2H, Q_{AB} , $J=5.2$ Hz, H-15 and H-16), 4.95 (1H, dd, $J_1=2.1$ Hz, $J_2=3.5$ Hz, H-7), 4.58 (1H, bs, H-14), 3.77 and 3.56 (2H, Q_{AB} , $J=11.2$ Hz, 2H-18), 2.94 (3H, s, $\text{CH}_3\text{-SO}_2$ -), 2.08 and 2.04 (3H each, s, AcO groups at C-18 and C-7); 1.15, 0.81 and 0.80 (3H, each, s, Me groups at C-17, C-19 and C-20); ^{13}C nmr (100 MHz, CDCl_3): see Table 1; ms, m/z(%): $[\text{M}+1]^+$ 483 (0.05), 423 (0.2), 387 (1.3), 363 (1.6), 327 (59.7), 267 (100). Anal. Found: C, 62.17; H, 8.02; S, 6.41. Calcd. for $\text{C}_{25}\text{H}_{38}\text{O}_7\text{S}$: C, 62.22; H, 7.94; S, 6.64.

Reduction of product 4

Product 4 (100 mg) was dissolved in THF (10 mL) and treated with LiAlH_4 (10 mg) at room temperature for 48 h. The reaction mixture was diluted with aqueous diethyl ether, extracted with CH_2Cl_2 , dried with anhydrous Na_2SO_4 and evaporated in vacuum to dryness. Chromatography over silica gel gave 48 mg of *ent*-7 α ,18-dihydroxy-12(13) \rightarrow 14-*abeobeyer*-13,15-diene (5, 48%); m.p.: 225°C ; $[\alpha]_{\text{D}} = 70.4^\circ$ (CHCl_3 , c 1); IR_{vmax} (CHCl_3): 3378, 1460, 754 cm^{-1} ; ^1H nmr (400 MHz, CDCl_3): δ 6.14 and 6.04 (2H, Q_{AB} , $J=5.7$ Hz, H-15 and H-16), 4.79 and 4.75 (1H each, s, 2H-17), 3.72 (1H, dd, $J_1=J_2=2.9$ Hz H-7), 3.46 and 2.96 (2H, Q_{AB} , $J=11.3$ Hz, 2H-18), 2.81 (1H, d, $J=11.1$ Hz, H-14), 0.90 and 0.73 (3H each, s, Me groups at C-19 and C-20); ^{13}C nmr (100 MHz, CDCl_3): see Table 1; ms, m/z(%): $[\text{M}+1]^+$ 303 (13), 285 (53), 267 (14), 57 (100). Anal. Found: C, 79.22; H, 10.08. Calcd. for $\text{C}_{20}\text{H}_{30}\text{O}_2$: C, 79.42; H, 10.00; and 35 mg of *ent*-7 α ,14 β ,18-trihydroxybeyer-15-ene (6, 35%)¹¹; IR_{vmax}

(CHCl₃): 3383, 1456, 750 cm⁻¹; ¹H nmr (400 MHz, CDCl₃): δ 5.49 and 5.47 (2H, Q_{AB}, J=6.6 Hz, H-15 and H-16), 4.02 (1H, dd, J₁=J₂=3.0 Hz, H-7), 3.50 and 2.92 (2H, Q_{AB}, J=11.4 Hz, 2H-18), 1.06, 0.76 and 0.70 (3H each, s, Me groups at C-17, C-19 and C-20); ¹³C nmr (100 MHz, CDCl₃): see Table 1; ms, m/z(%): [M+1]⁺ 321 (1), 303 (25), 285 (100), 267 (36).

Solvolysis of product 4: Pathway A: Acetolysis

Mesylate 4 (100 mg) was treated with AcOK/AcOH 0.5 N (4 mL). The reaction mixture was refluxed for 15 min and then neutralised with NaHCO₃, extracted with CH₂Cl₂, dried over anhydrous Na₂SO₄ and evaporated in vacuum to dryness. The crude product was chromatographed on a silica gel column to give 95 mg of *ent*-7α,18-diacetoxy-12(13)→14-*abeobeyer*-13,15-diene (7, 95%); m.p.: 93-95 °C; [α]_D = 72.9° (CHCl₃, c 1); IR_{vmax} (CHCl₃): 3076, 1739, 1635, 1247, 1039 and 757 cm⁻¹; ¹H nmr (400 MHz, CDCl₃): δ 6.18 and 6.00 (2H, Q_{AB}, J=5.7 Hz, H-15 and H-16), 4.80 and 4.71 (1H each, s, 2H-17), 4.80 (1H, bs, H-7), 4.28 and 3.65 (2H, Q_{AB}, J=11.1 Hz, 2H-18), 2.60 (1H, bd, J=10.8 Hz, H-14), 2.07 and 2.03 (3H each, s, AcO groups at C-18 and C-7), 0.91 and 0.84 (3H each, s, Me groups at C-19 and C-20); ¹³C nmr (100 MHz, CDCl₃): see Table 1; ms, m/z(%): [M+1]⁺ 387 (1), 327 (21), 269 (100), 267 (49). Anal. Found: C, 74.32; H, 8.94. Calcd. for C₂₄H₃₄O₄: C, 74.58; H, 8.87.

Solvolysis of product 4: Pathway B: Treatment with EtOH/Zn

Zinc dust (100 mg) was added to a solution of 100 mg of product 4 in 10 mL EtOH, then the reaction mixture was heated to reflux for 12 h, and the solvent was removed in vacuum. The residue was extracted with CH₂Cl₂, and the organic layer was washed with NaHCO₃ solution, dried over anhydrous sodium sulphate and evaporated in vacuum. Chromatography on a silica gel column yielded 90 mg of the product 7 (90%).

Acetylation of product 5

Product 5 (20 mg) was dissolved in Ac₂O/Py (0.5:1 ml) and refluxed for 12 h. The reaction mixture was diluted with water, extracted with CH₂Cl₂, washed with saturated KHSO₄ and dried with anhydrous Na₂SO₄. Chromatography over silica gel yielded 18 mg of diacetate 7 (72%).

Acid treatment of product 7

A very small amount (5 mg) of TsOH (p-toluenesulfonic acid) was added to a solution of product 7 (50 mg) in toluene (5 mL) and the reaction mixture was heated at 140°C for 12 h in a sealed tube. The reaction mixture was cooled to room temperature, evaporated in vacuum and chromatographed on a silica gel column. This yielded 35 mg of starting product and 11 mg of *ent*-18-acetoxy-12(13)→14,15(8)→7-*diabeobeyer*-7,13,15-triene (15, 21%); m.p.: syrup; [α]_D = 5° (CHCl₃, c 1); IR_{vmax} (CHCl₃): 1741, 1454, 1237 and 804 cm⁻¹; ¹H nmr (400 MHz, CDCl₃): δ 6.90 and 6.84 (2H, Q_{AB}, J=7.5 Hz, H-15 and H-16), 3.93 and 3.74 (2H, Q_{AB}, J=11.0 Hz, 2H-18), 2.23 (3H, s, Me group at C-17), 2.06 (3H, s, AcO groups at C-18), 1.00 and 0.88 (3H each, s, Me groups at C-19 and C-20), ¹³C nmr (100 MHz, CDCl₃): see Table 1; ms, m/z(%): [M+1]⁺ 327 (2), 297 (4), 267 (36), 51 (100). Anal. Found: C, 80.56; H, 9.54. Calcd. for C₂₂H₃₀O₂: C, 80.94; H, 9.26.

Hydrogenation of product 2

Product 2 (150 mg) was hydrogenated according to reference 14 to give 135 mg of *ent*-7α,18-diacetoxy-14β-hydroxybeyerane (8, 90%)¹⁴.

Mesylation of product 8

Methanesulfonyl chloride (0.2 mL) was added to a solution of 120 mg of product 8 dissolved in 0.4 mL pyridine. The reaction mixture was stirred at room temperature for 1 h. Then the reaction mixture was diluted with CH₂Cl₂, washed with water and with saturated aqueous KHSO₄, and concentrated in

vacuum. Chromatography on a silica gel column yielded 114 mg of *ent-7 α ,18*-diacetoxy-14 β -mesyloxybeyerane (**9**, 95%); m.p.: 140-2°C; $[\alpha]_D = 82.7^\circ$ (CHCl₃, c 1); IR_{vmax} (CHCl₃): 1732, 1335, 1250, 1172 cm⁻¹; ¹H nmr (400 MHz, CDCl₃): 4.81 (1H, dd, J₁=1.9 Hz, J₂=3.9 Hz, H-7), 4.61 (1H, bs, H-14), 3.78 and 3.54 (2H, Q_{AB}, J=11.2 Hz, 2H-18), 2.99 (3H, s, CH₃-SO₂-), 2.08 and 2.04 (3H each, s, AcO groups at C-18 and C-7), 1.10, 0.99 and 0.80 (3H, each, s, Me groups at C-17, C-19 and C-20); ¹³C nmr (100 MHz, CDCl₃): see Table 1; ms, m/z(%): [M+1]⁺ 485 (0.3), 425 (0.2), 365 (1.6), 330 (7.8), 329 (31.8), 269 (100). Anal. Found: C, 61.65; H, 8.54; S, 6.27. Calcd. for C₂₅H₄₀O₇S: C, 61.96; H, 8.32; S, 6.62.

Acetolysis of product 9

Mesyate **9** (100 mg) was treated with AcOK/AcOH 0.5 N (4 mL). The reaction mixture was refluxed for 15 min and then neutralised with NaHCO₃, extracted with CH₂Cl₂, dried over anhydrous Na₂SO₄ and evaporated in vacuum to dryness. The crude product was chromatographed on a silica gel column to give 95 mg of *ent-7 α ,18*-diacetoxy-12(13)→14-*abeobeyer*-16-ene (**10**, 95%); m.p.: 116-118°C; $[\alpha]_D = 57.3^\circ$ (CHCl₃, c 1); IR_{vmax} (CHCl₃): 1738, 1247; ¹H nmr (400 MHz, CDCl₃): δ 5.04 (1H, bs, H-16), 4.82 (1H, dd, J₁=J₂=2.5 Hz, H-7), 3.70 and 3.62 (2H, Q_{AB}, J=11.1 Hz, 2H-18), 2.48 (1H, dd, J₁=16.4 Hz, J₂=4.6 Hz, H-15), 2.28 (1H, bd, J=10.4 Hz, H-14), 2.05 and 2.03 (3H each, s, AcO groups at C-18 and C-7), 1.60 (3H, bs, Me group at C-17), 0.83 and 0.80 (3H each, s, Me groups at C-19 and C-20); ¹³C nmr (100 MHz, CDCl₃): see Table 1; ms, m/z(%): [M+1]⁺ 389 (21), 329 (21), 269 (100). Anal. Found: C, 73.89; H, 9.02. Calcd. for C₂₄H₃₆O₄: C, 74.19; H, 9.34.

Oxidation of product 2

Product **2** (200 mg) was oxidized by the Jones method according to reference 14, and 190 mg of *ent-7 α ,18*-diacetoxy-14-oxo-beyer-15-ene (**11**, 95%)¹⁴ were obtained.

Reduction of product 11

NaBH₄ (15 mg) was added to a solution of product **11** (150 mg) in MeOH (10 mL) and the mixture was stirred at room temperature for 1 h. The reaction mixture was acidified with dil. HCl, extracted with CH₂Cl₂, dried with anhydrous Na₂SO₄ and concentrated in vacuum. Chromatography on a silica gel column yielded 135 mg of *ent-7 α ,18*-diacetoxy-14 α -hydroxybeyer-15-ene (**12**, 90%) m.p.: 162-4°C; $[\alpha]_D = 12.6^\circ$ (CHCl₃, c 1); IR_{vmax} (CHCl₃): 3590, 1737, 1248 cm⁻¹; ¹H nmr (400 MHz, CDCl₃): δ 5.58 and 5.49 (2H, Q_{AB}, J=5.2 Hz, H-15 and H-16), 4.90 (1H, dd, J₁=2.2 Hz, J₂=3.4 Hz, H-7), 3.70 and 3.65 (2H, Q_{AB}, J=11.0 Hz, 2H-18), 3.35 (1H, bs, H-14), 2.07 and 2.04 (3H each, s, AcO groups at C-18 and C-7), 0.98 (3H, s, Me group at C-17), 0.82 (6H, s, Me groups at C-19 and C-20); ¹³C nmr (100 MHz, CDCl₃): see Table 1; ms, m/z(%): [M]⁺ 404 (0.5), 344 (16.6), 284 (11.0), 269 (15.0), 43 (100). Anal. Found: C, 71.20; H, 8.86. Calcd. for C₂₄H₃₆O₅: C, 71.26; H, 8.97.

Mesylation of product 12

Methanesulfonyl chloride (0.2 mL) was added to a solution of 120 mg of product **12** dissolved in 0.4 mL of pyridine. The reaction mixture was stirred at room temperature for 1 h. Then, the reaction mixture was diluted with CH₂Cl₂, washed with water and with saturated aqueous KHSO₄ and concentrated in vacuum. Chromatography on a silica gel column yielded 114 mg of *ent-7 α ,18*-diacetoxy-14 α -mesyloxybeyerane (**13**, 95%); m.p.: 145-7°C; $[\alpha]_D = 56^\circ$ (CHCl₃, c 1); IR_{vmax} (CHCl₃): 1732, 1362, 1246, 1174 cm⁻¹; ¹H nmr (400 MHz, CDCl₃): δ 5.56 (2H, collap. Q_{AB}, H-15 and H-16), 4.89 (1H, dd, J₁=J₂=2.5 Hz, H-7), 4.41 (1H, bs, H-14), 3.69 and 3.62 (2H, Q_{AB}, J=11.1 Hz, 2H-18), 3.07 (3H, s, CH₃-SO₂-), 2.11 and 2.04 (3H each, s, AcO groups at C-18 and C-7), 1.08, 0.83 and 0.82 (3H, each, s, Me groups at C-17, C-19 and C-20); ¹³C nmr (100 MHz, CDCl₃): see Table 1; ms, m/z(%): [M]⁺ 482 (0.1), 422 (0.2), 387 (0.3), 362 (0.6), 327 (1.7), 267 (1.6), 43 (100). Anal. Found: C, 61.85; H, 8.04; S, 6.32. Calcd. for C₂₅H₃₈O₇S: C, 62.22; H, 7.94; S, 6.64.

Acetolysis of product 13

Mesylate 13 (100 mg) was treated with AcOK/AcOH 0.5 N (4 mL). The reaction mixture was refluxed for 6 h and then neutralised with NaHCO₃, extracted with CH₂Cl₂, dried over anhydrous Na₂SO₄ and evaporated in vacuum to dryness. The crude product was chromatographed on a silica gel column to give a very difficult separation mixture of two products. This mixture was saponified with alcoholic KOH at room temperature for 24 hours to yield 90 mg of *ent*-7 α ,14 α ,18-trihydroxybeyer-15-ene (14, 90%); m.p.: 177-79 °C; [α]_D = +41.2° (CHCl₃, c 1); IR_{vmax} (CHCl₃): 3351, 3051 cm⁻¹; ¹H nmr (400 MHz, CDCl₃): δ 5.53 and 5.48 (2H, Q_{AB}, J=6.3 Hz, H-15 and H-16), 3.83 (1H, dd, J₁=J₂=2.7 Hz, H-7), 3.45 (1H, bs, H-14), 3.44 and 2.93 (2H, Q_{AB}, J=11.1 Hz, 2H-18), 0.98, 0.79 and 0.70 (3H, each, s, Me groups at C-17, C-19 and C-20); ¹³C nmr (100 MHz, CDCl₃): see Table 1; ms, m/z(%): [M]⁺ 320 (0.1), 302 (15.0), 284 (8.0), 43 (100). Anal. Found: C, 74.82; H, 9.91. Calcd. for C₂₀H₃₂O₃: C, 74.96; H, 10.06.

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REFERENCES AND NOTES

1. a) McAlees, A.J., McCrindle, R., *J. Chem. Soc. Perkin I*, **1975**, 861.
b) McAlees, A.J., McCrindle, R., Murphy, S.T., *J. Chem. Soc. Perkin I*, **1976**, 1042
2. a) Coates, R.M., Bertram, E.F., *J. Org. Chem.*, **1971**, *24*, 3722.
b) McAlees, A.J., McCrindle, R., Murphy, S.T., *J. Chem. Soc. Perkin I*, **1975**, 1641.
3. a) Yoshikoshi, A., Kitadani, M., Kitahara, Y., *Tetrahedron*, **1967**, *23*, 1175.
b) Hanson, J.R., *Tetrahedron*, **1967**, *23*, 793.
c) Rodríguez, B., Valverde, S., *Chem. Ind.*, **1976**, 1010.
4. Coates, R.M., "Biogenetic-Type Rearrangements of Terpenes" in *Prog. Chem. Org. Nat. Prod.*, **1976**, *33*, 73.
5. Murray, R.D., Mills, R., W., McAlees, A.J., McCrindle, R., *Tetrahedron*, **1974**, *30*, 3399.
6. Yun-Xing Cheng, Wei-Shan Zhou, Hou-Ming Wu, *Tetrahedron*, **1993**, *49*, 97.
7. García-Granados, A., Parra, A., *Tetrahedron*, **1991**, *47*, 9103.
8. García-Granados, A., Martínez, A., Onorato, M.E., *J. Org. Chem.*, **1987**, *52*, 606.
9. García-Granados, A., Parra, A., Peña, A., Socorro, O., *An. Quim.*, **1984**, *80*, 175.
10. García-Granados, A., Martínez, A., Onorato, M.E., Arias, J.M., *J. Nat. Prod.*, **1985**, *48*, 371.
11. García-Granados, A., Martínez, A., Onorato, M.E., Socorro, O., *Phytochem.*, **1984**, *23*, 607.
12. Llinas, J.R., Vincent, E.J., Peiffer, G., *Bull. Soc. Chim. France*, **1973**, *11*, 3208.
13. Program P.C. MODEL available from SERENA SOFTWARE, P.O. BOX 3076, BLOOMINGTON, I.N. 47402-3076, U.S.A.
14. García-Granados, A., Onorato, M.E., Santoro, J., *Mag. Res. Chem.*, **1986**, *24*, 853

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